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(54) **ANCHORS WITH OPEN HEADS**

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See application file for complete search history.

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Sep. 11, 2009.

(51) **Int. Cl.**

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(Continued)

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CPC **A61F 2/04** (2013.01); **A61F 5/0076**
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(2013.01);

(Continued)

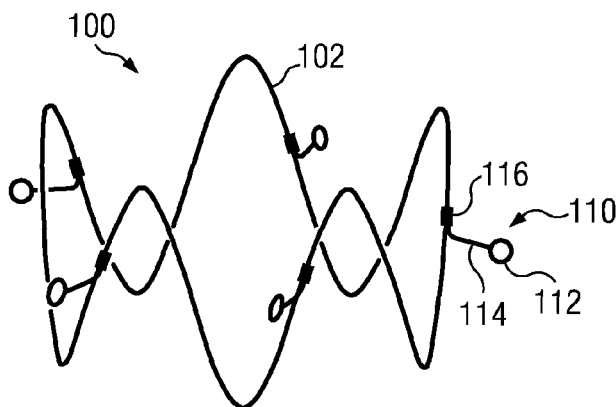
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CPC A61F 2/2409; A61F 2/04; A61F 2/848;
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A61F 2220/0016; A61F 2230/0091

ABSTRACT

An implant (100) includes a protrusion (110) with an open or rounded loop (112) (or open head) connected to a collapsible anchor. The protrusion may include a straight length of wire (114) or a helical length wire, with one or more wire loops (112) at the end of the length forming the loop. Upon deployment within the gastrointestinal tract, the protrusion expands from a collapsed state, alongside the anchor, to a relaxed state, in which the protrusion extends outward from the anchor. As the protrusion expands to its relaxed state, it pushes the loop into the wall of the duodenum, causing the loop to penetrate the duodenal wall. A pocket of scar tissue forms about the head and possibly through an opening in the head, securing the anchor within the duodenum. The implant may also include a thin-walled sleeve that is coupled to the anchor and extended from the anchor into the intestine.

36 Claims, 10 Drawing Sheets



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CPC *A61F 5/0089* (2013.01); *A61F 2002/045*
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2220/0008 (2013.01); *A61F 2220/0016*
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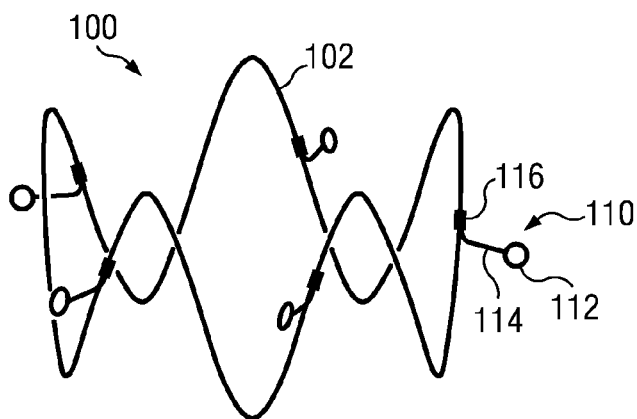


FIG. 1A

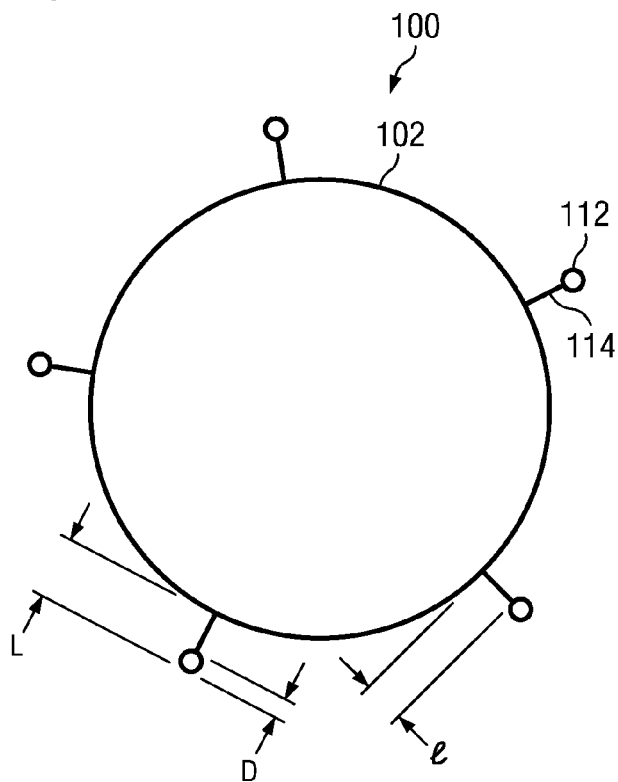


FIG. 1B

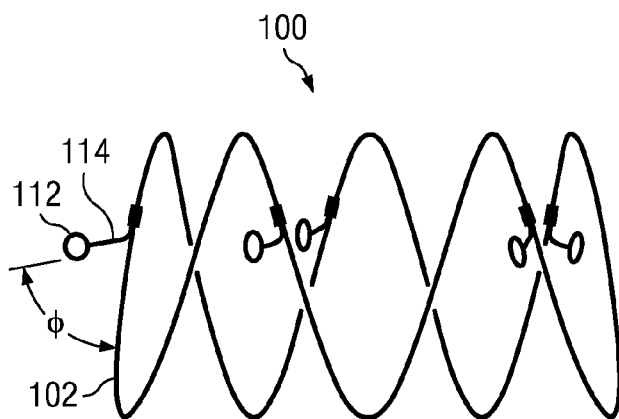


FIG. 1C

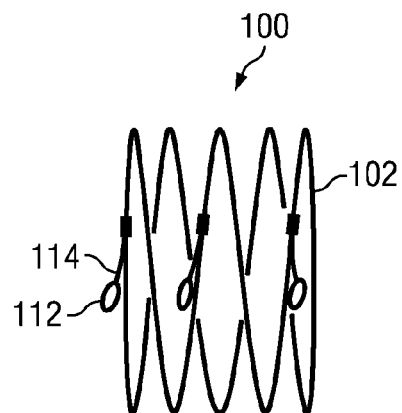
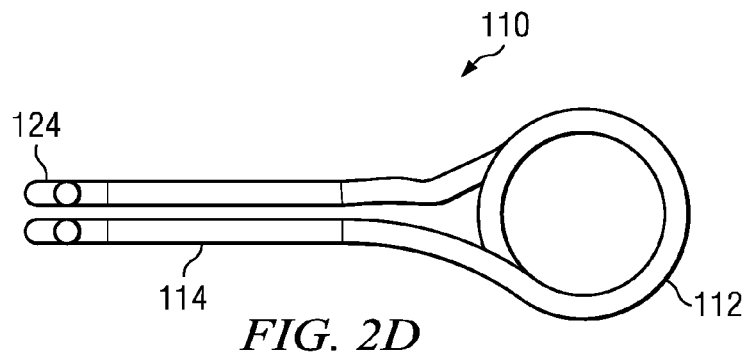
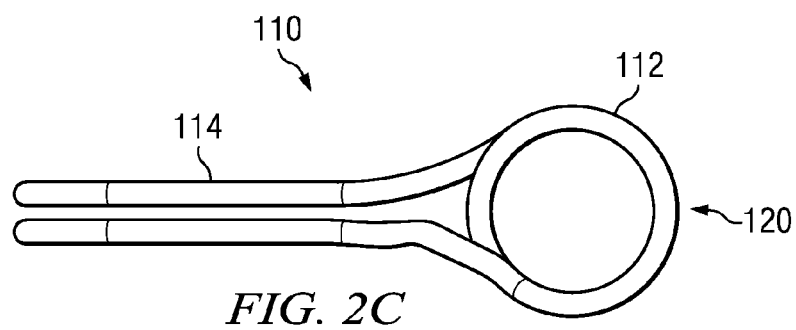
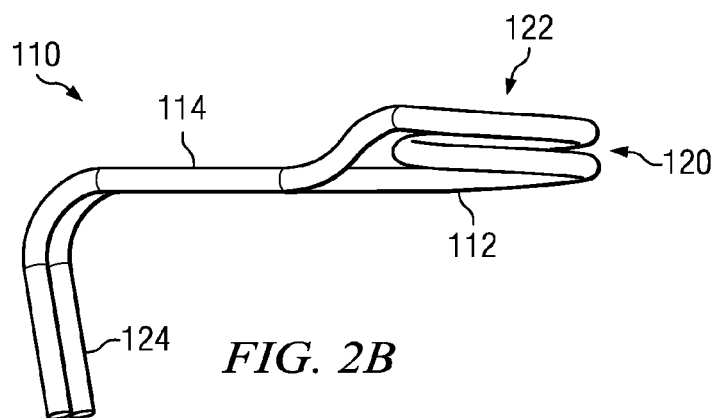
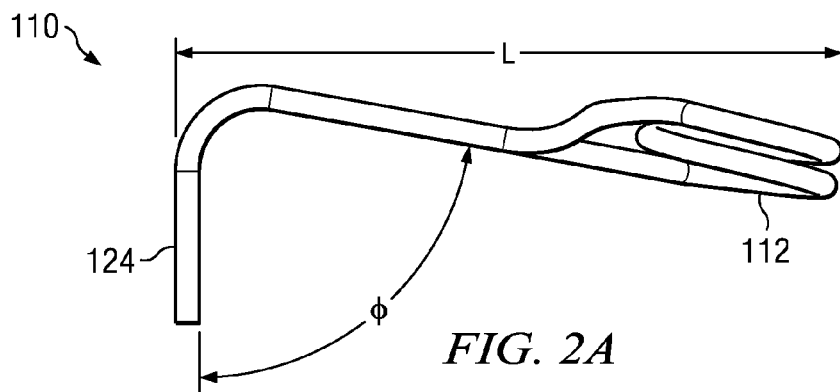


FIG. 1D



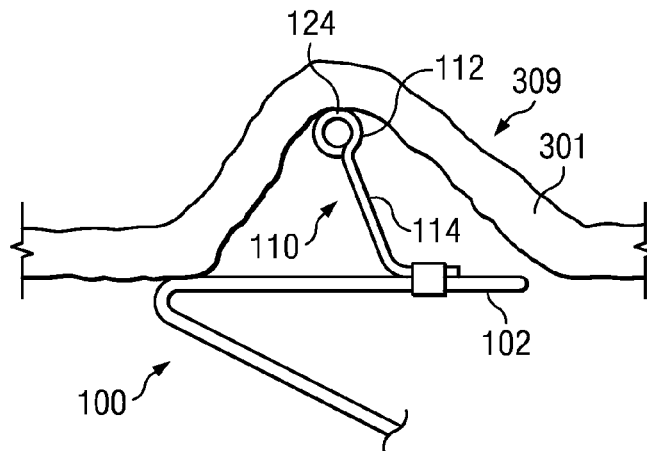


FIG. 3A

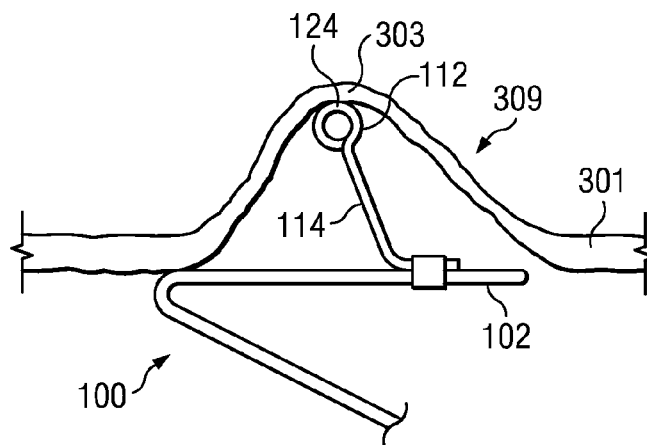


FIG. 3B

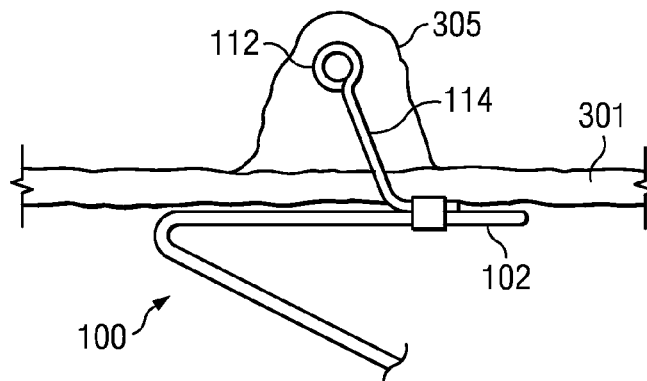


FIG. 3C

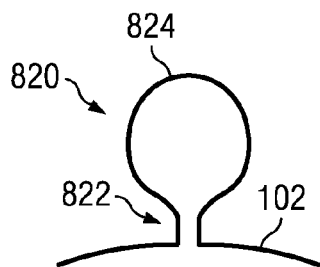


FIG. 4A

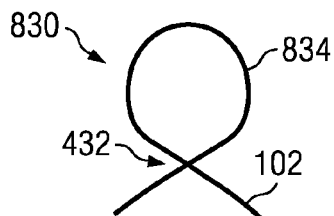


FIG. 4B

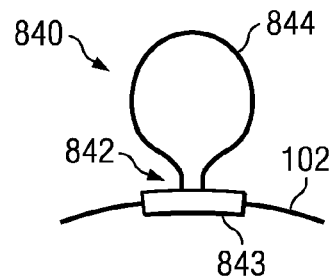


FIG. 4C

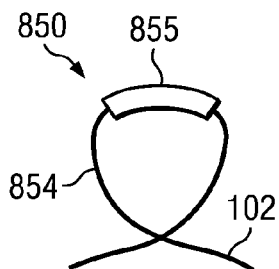


FIG. 4D

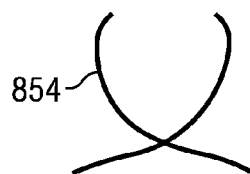


FIG. 4E

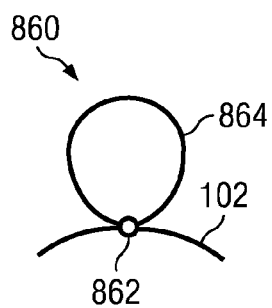


FIG. 4F

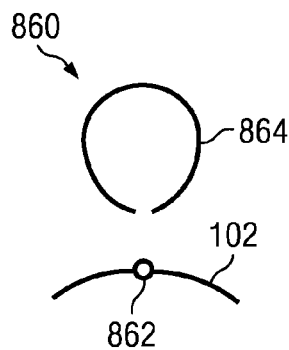


FIG. 4G

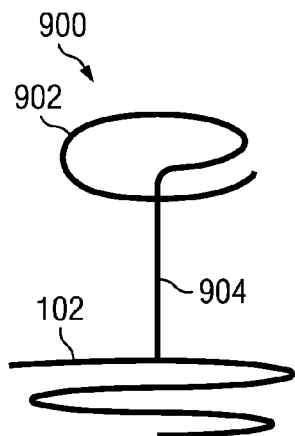


FIG. 4H

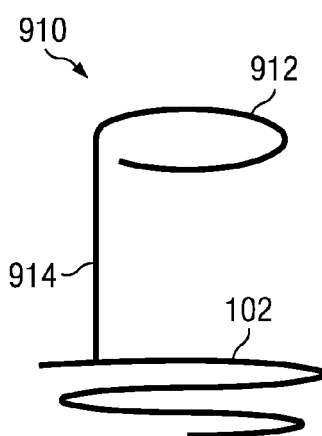


FIG. 4I

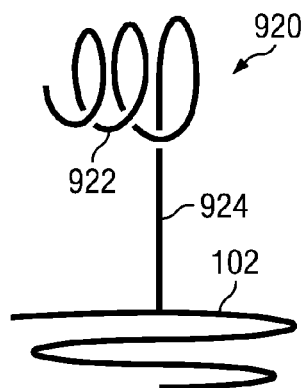


FIG. 4J

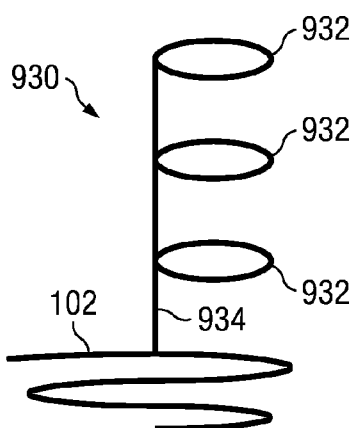


FIG. 4K

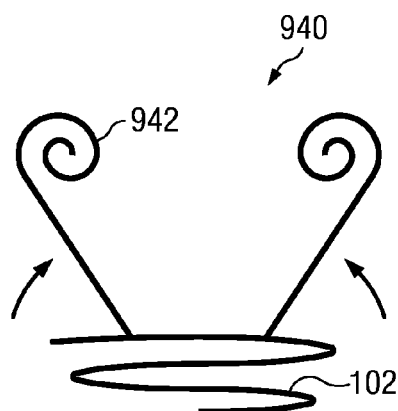


FIG. 4L

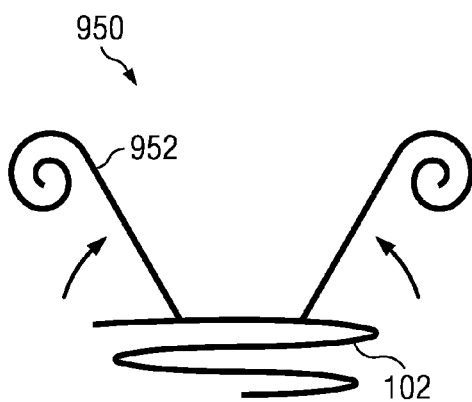


FIG. 4M

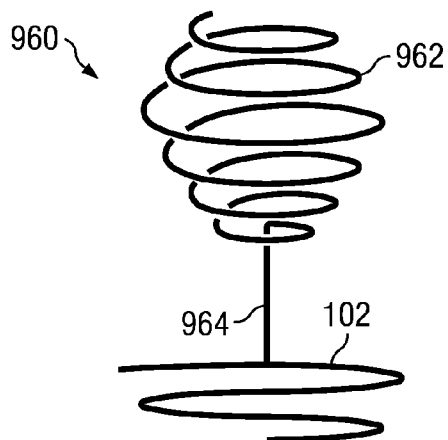


FIG. 4N

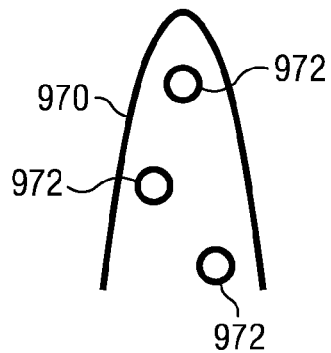


FIG. 4O

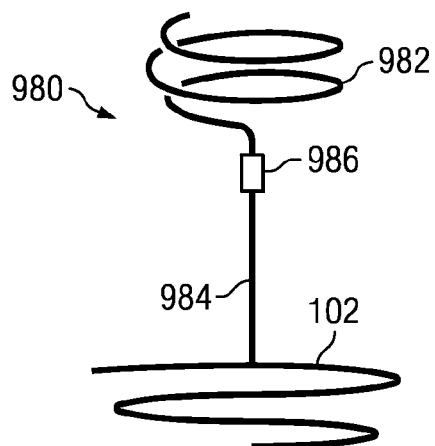


FIG. 4P

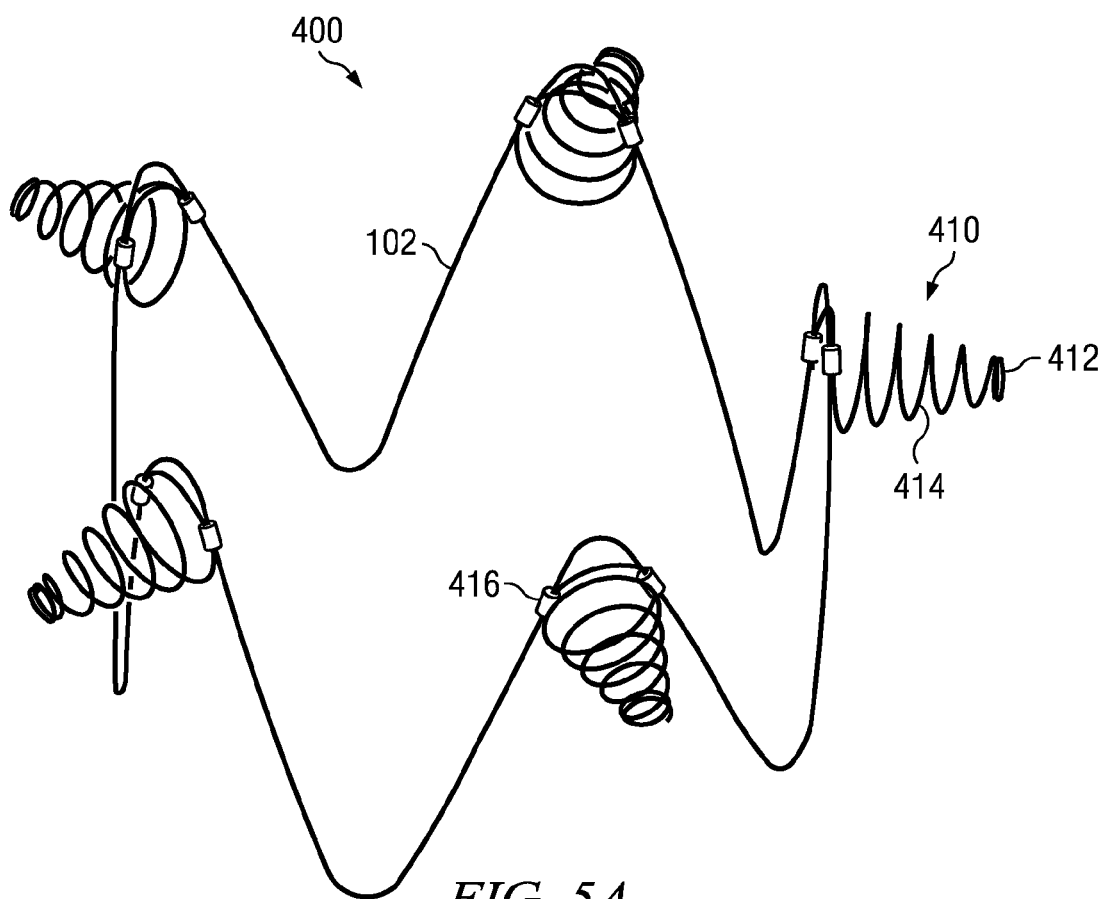
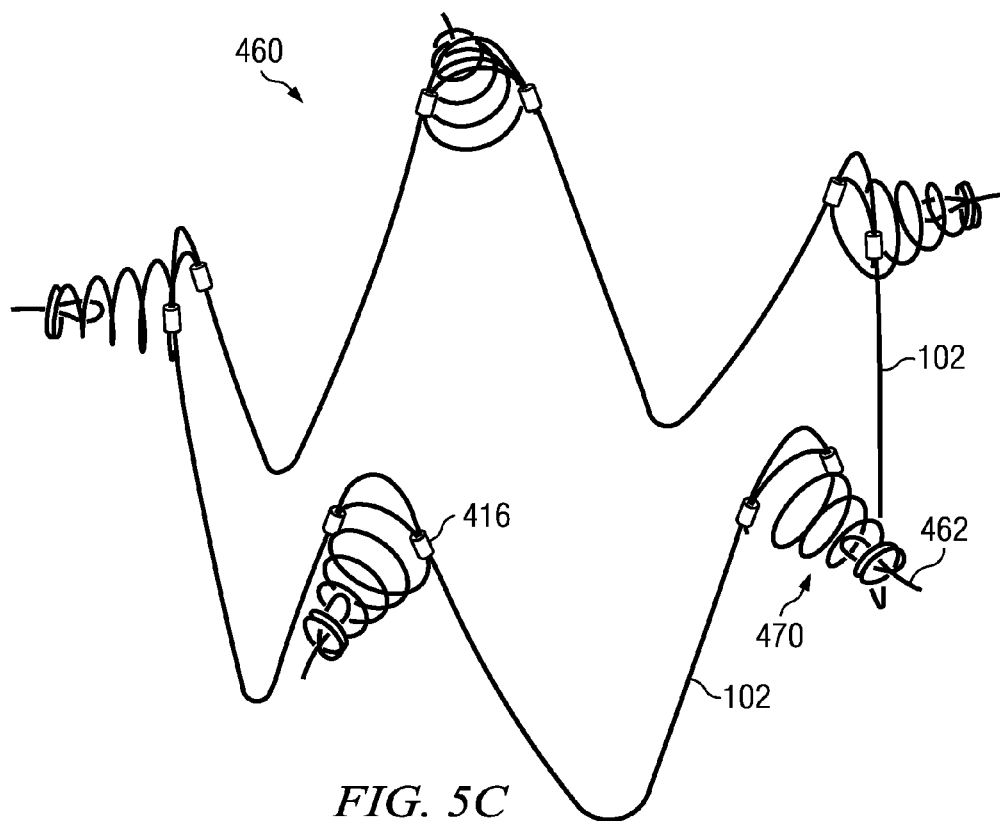
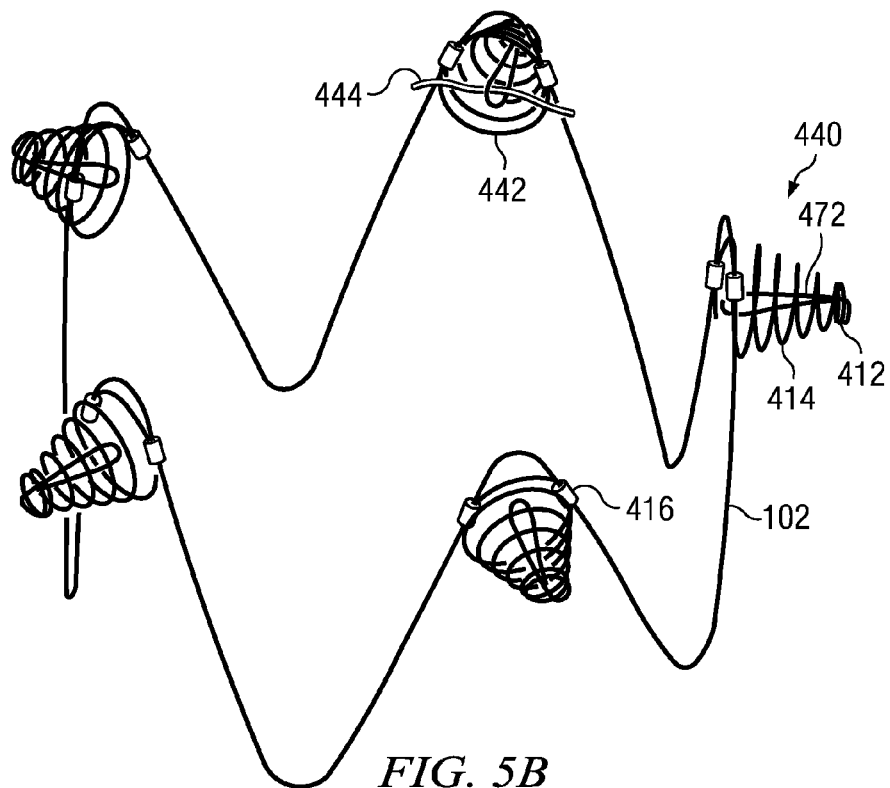


FIG. 5A



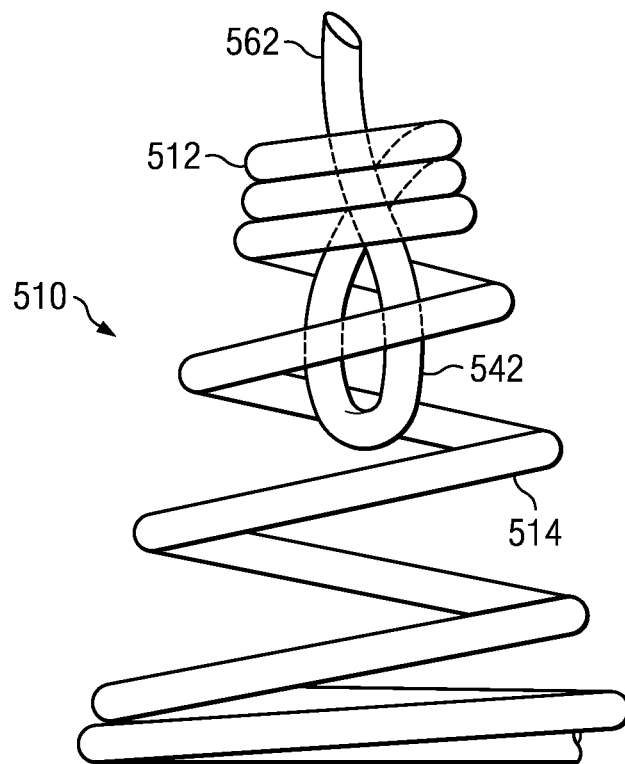
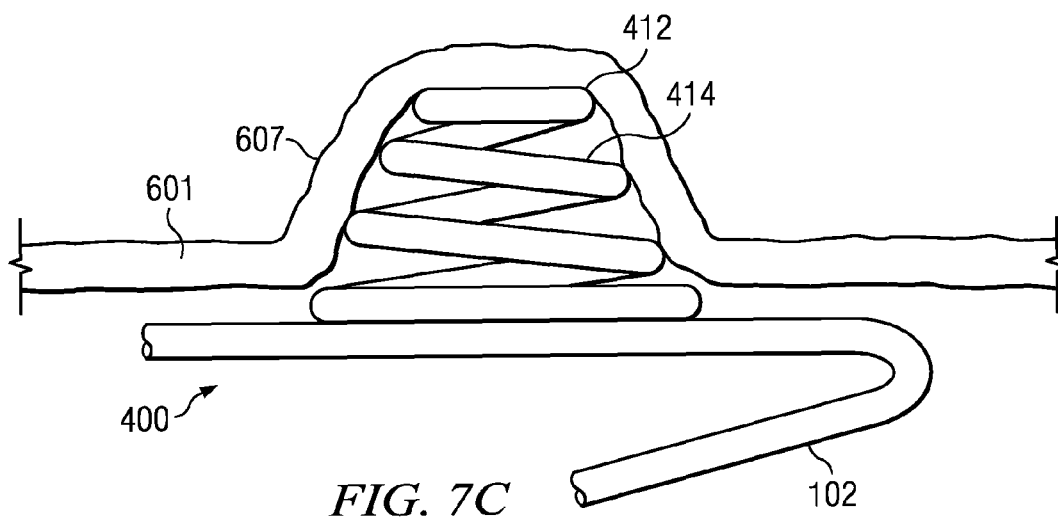
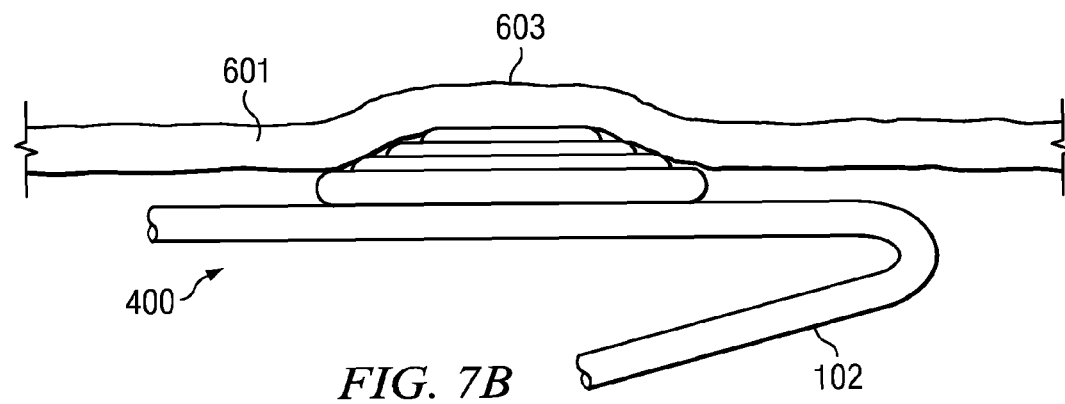
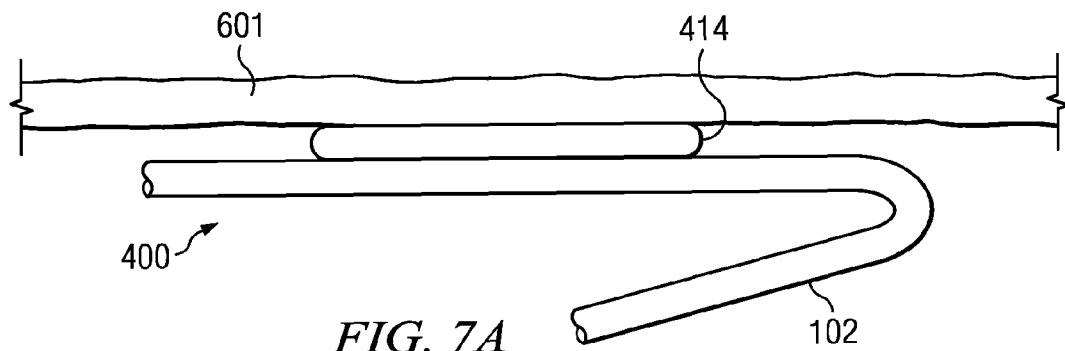
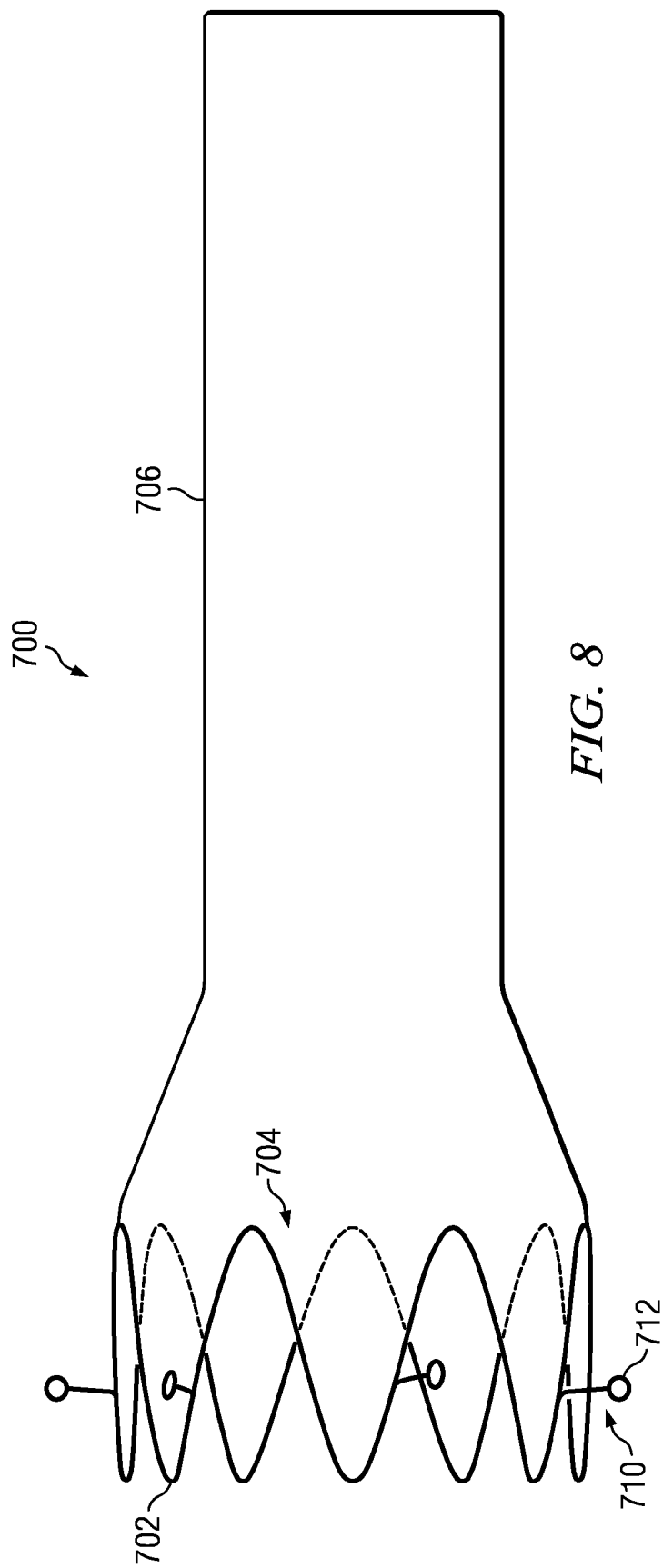


FIG. 6





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ANCHORS WITH OPEN HEADS**RELATED APPLICATIONS**

This application is the U.S. National Stage of International Application No. PCT/US2010/048444, filed Sep. 10, 2010, which designates the U.S., published in English, and claims the benefit of U.S. Provisional Application Nos. 61/361,806 filed Jul. 6, 2010, and 61/276,381 filed Sep. 11, 2009. The entire teachings of the above applications are incorporated herein by reference.

BACKGROUND

Obesity is associated with a wide variety of health problems, including Type 2 diabetes, hypertension, coronary artery disease, hypercholesterolemia, sleep apnea, and pulmonary hypertension. It also exerts an enormous strain on the body that affects the organs, the nervous system, and the circulatory systems. Obesity rates have been rising for years in the United States, causing corresponding increases in healthcare expenditures.

Curing obesity has so far vexed the best efforts of medical science. Dieting is not an adequate long-term solution for most obese people, especially those with a body-mass index of over 30. Stomach stapling, or gastropasty, reduces the size of the stomach, leading to reduced appetite and weight loss, but eventually the stomach stretches and the patient's appetite returns to pre-surgery levels. Roux-en-Y gastric bypass reduces the size of the stomach and the length of the intestine, and leads to both weight loss and alleviation of the Type 2 diabetes common to obese patients. Although gastric bypass appears to provide a more permanent solution than gastropasty, complication rates associated with gastric bypass are between 2% and 6%, with mortality rates of about 0.5-1.5%.

Endoscopically delivered gastrointestinal implants, such as those described in commonly assigned U.S. Pat. Nos. 7,025,791 and 7,608,114 to Levine et al., incorporated herein by reference in their entireties, provide the benefits of gastric bypass without the hazards of surgery. For example, an implant may include a thin-walled, floppy sleeve that is secured in the stomach or intestine with a collapsible anchor. The sleeve extends into the intestine and channels partially digested food, or chyme, from the stomach through the intestine in a manner that may cause weight loss and improve diabetes symptoms. The sleeve and anchor can be removed endoscopically when treatment is over or if the patient desires.

SUMMARY

Embodiments of the present invention provide improved anchoring of an implant in the gastrointestinal tract and can increase the duration that an implant can be anchored in the intestine by encouraging stable tissue reactions to the implant. An implant according to principles of the present invention may include a long protrusion with an open loop connected to a collapsible anchor. The collapsible anchor, which may, for example, be a wave anchor or a stent comprising a network of struts, is configured to be deployed within a lumen in a mammalian body. Upon deployment, the collapsible anchor expands within the lumen, and the protrusion expands away from the anchor, pushing the open loop against the wall of the lumen. Over time, the protrusion and the open loop penetrate the luminal wall, and the open loop may project through the far side of the luminal wall. A pocket of scar tissue forms about the open loop and through an

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opening in the open loop, securing the anchor within the lumen. The implant may have additional protrusions, each of which is connected to the anchor and includes an open loop. Each additional open loop also includes an opening and is adapted to penetrate the luminal wall upon deployment of the collapsible anchor.

Each open loop may have an inner opening with a width of between about 1 mm and about 13 mm, or, more preferably, an inner diameter of about 3 mm. Typically, the protrusion extends along a total length of between about 6 mm and about 13 mm from the collapsible anchor upon full deployment from the collapsible anchor. The protrusion and the open loop may be formed of wire (e.g., nitinol wire) with a preferred diameter of about 0.010 inch to about 0.040 inch, and more preferably about 0.020 inch.

The open loop can be formed of a loop of wire, and the protrusion can be formed of a straight length of wire extending from the loop of wire. The open loop may be oriented in a variety of directions with respect to the collapsible anchor. For example, the open loop may define a plane that is perpendicular to the lumen wall when the protrusion is deployed. Alternatively, the open loop may define a plane that is parallel to the lumen wall when the protrusion is deployed. When the protrusion is in a collapsed state, it folds against or along the side of the collapsible anchor. When relaxed, straight protrusions typically extend outwards from the collapsible anchor at an angle of between about 45 degrees and about 135 degrees, or, more preferably, to an angle of about 80 degrees or about 90 degrees. At these angles, the expanded straight protrusion pushes the loop outward, causing an edge of the loop to engage the luminal wall.

Alternatively, the protrusion can include a length of wire formed in a helix. The wire used to form the helix may be coiled to form the loop, which can be oriented such that it is parallel to the luminal wall when deployed within the lumen. (Other orientations of the loop are also possible.) The helix may have a tapered profile (e.g., a conical profile) when viewed from the side, and can be flattened alongside the collapsible anchor. The collapsed implant can be inserted into the lumen endoscopically. Releasing the helix and the anchor from the collapsed state causes the helix to push the loop away from the anchor, which, in turn, causes a face of the loop to engage the luminal wall. The implant may also include an end effect at or near the tip of the loop to aid penetration of the loop through the luminal wall.

The implant can be collapsed, for removal from the lumen, with an optional drawstring that runs through the opening in the loop or through additional retaining hooks or loops connected to the loop or the protrusion. Pulling on the drawstring collapses the loop and protrusion towards the collapsible anchor, and away from the luminal wall. Collapsing an implanted helix may cause coils in the helix to shear fibrotic tissue formed about the helix depending on the spacing and orientation of the coils that make up the helix.

An implant with a protrusion and an open loop can also include an unsupported, thin-walled sleeve coupled to the collapsible anchor and configured to extend into the lumen (e.g., the intestine) upon deployment of the collapsible anchor. The implant may also include a restrictor plate instead of or in addition to the thin-walled sleeve.

BRIEF DESCRIPTION OF THE DRAWINGS

The foregoing will be apparent from the following more particular description of example embodiments of the invention, as illustrated in the accompanying drawings in which like reference characters refer to the same parts throughout

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the different views. The drawings are not necessarily to scale, emphasis instead being placed upon illustrating embodiments of the present invention.

FIGS. 1A-1D are perspective, plan, and elevation views of straight protrusions with open loops coupled to a wave anchor.

FIGS. 2A-2D are plan and elevation views of a straight protrusion with an open loop suitable for connection to an anchor.

FIGS. 3A-3C illustrate how a straight protrusion with an open loop penetrates the wall of the gastrointestinal tract and how a fibrotic encapsulation forms about and through the open loop.

FIGS. 4A-4P show various alternative protrusions with open loops.

FIGS. 5A-5C are perspective views of wave anchors with helical protrusions with open loops.

FIG. 6 is a perspective view of a loop projection with a helical neck.

FIGS. 7A-7C illustrate how a helical protrusion with an open loop penetrates the wall of the gastrointestinal tract and how a fibrotic encapsulation forms about and through the helical protrusion and the open loop.

FIG. 8 shows an implant that includes a sleeve extending from an anchor with open-tip protrusions.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

A description of example embodiments of the invention follows. An anchor may be used to secure a sleeve in the intestine of a patient for treating obesity and/or type-2 diabetes as described in commonly assigned U.S. Pat. Nos. 7,025, 791; 7,608,114; 7,476,256; U.S. patent application Ser. No. 10/858,852, filed on Jun. 1, 2004, by Levine et al.; U.S. patent application Ser. No. 11/330,705, filed on Jan. 11, 2006, by Levine et al.; U.S. patent application Ser. No. 11/827,674 filed on Jul. 12, 2007, by Levine et al., all of which are incorporated herein by reference in their entireties.

As described in the above-referenced patents and patent applications, straight, sharp barbs fixed to a self-expanding anchor may be used to secure an implant to the duodenal wall. However, the body's healing response stimulates a progressive tissue proliferation around sharp barbs in response to the injury caused as the anchor pushes the sharp barbs into the wall of the duodenum. The inflammatory response to the injury produces a mix of granulation and more stable fibrous tissue (i.e., scar tissue). This causes thickening of the duodenal wall over time resulting in barbs disengaging from the tissue. As sharp barbs separate from the duodenal wall, the implant may become unstable and migrate or rotate within the duodenum.

Long barbs tend to be better than short barbs at holding implants securely for longer periods. Without subscribing to any particular theory, it appears that longer barbs are more stable because it takes more time for the inflammatory thickening to separate longer barbs from the muscle layer. However, there is a practical limit to how long sharp barbs can be because longer sharp barbs are more likely to infiltrate surrounding organs. Very long sharp barbs can pierce through the muscle wall of the intestine and into adjacent structures and could potentially cause leaks, bleeding, or adhesions to other organs.

Protrusions with open loops (also called open heads), on the other hand, can secure an implant for longer periods of time while minimizing the risk of damage to nearby organs. In one embodiment, the protrusion, which is relatively narrow

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(e.g., about 0.060 inch wide) and relatively long (e.g., about 13 mm long), connects a relatively broad open loop (e.g., about 3 mm in diameter) to a collapsible anchor. Upon deployment, the protrusion pushes the open loop against the intestinal wall. Without being bound by any particular theory, initial research suggests that the muscle layer in the intestine stretches across the loop, and it eventually thins out or erodes enough to allow the loop to penetrate the luminal wall. A chronic inflammation response causes scar tissue to form around the loop and through the opening formed by the loop; this scar tissue can hold the loop securely. Because the loop is rounded or otherwise shaped to promote erosion through the muscle wall, the protrusion and the loop are less likely to pierce the scar tissue or surrounding organs.

Straight Protrusions with Open Loops

FIGS. 1A-1D show an implant **100** suitable for deployment within the gastrointestinal tract distal to the pylorus. FIGS. 1A-1C show perspective, plan, and elevation views of the implant **100** in a relaxed state (from the top, the relaxed implant **100** looks circular); FIG. 1D is an elevation view of the implant **100** in a compressed state. Typically, the implant **100** is compressed for endoscopic deployment within the gastrointestinal tract. Once positioned properly within the gastrointestinal tract, the implant **100** expands to the relaxed state shown in FIGS. 1A-1C.

The implant **100** includes a collapsible wave anchor **102** that includes a plurality of protrusions **110**, each of which extends outward from the wave anchor **102** when the implant **100** is in a relaxed state. The anchor **102** may have a relaxed diameter of about 40 mm or greater, e.g., about 45 mm, about 50 mm, or about 55 mm. Each protrusion **110** includes a rounded loop **112** at the end of a narrow, straight neck **114**, and each loop **112** includes an opening whose inner width **D** is within the range (inclusive) of between about 1 mm and about 13 mm, and preferably a diameter **D** within a range of about 1 mm and about 6 mm, or, more preferably, about 3 mm. The outer diameter can be within a range of about 2 mm to about 8 mm, and the diameter of the wire used to form each protrusion **110** can be within a range of about 0.010 inch to about 0.030 inch. Typically, the minimum bend radius of the wire limits the minimum inner diameter (it can be difficult to bend the wire too tightly), and the minimum desired pressure exerted by the loop **112** against the tissue limits the maximum inner diameter (bigger loops **112** may not exert enough pressure on the tissue to penetrate the tissue). The straight neck **114** has a length **l** of between about 6 mm and about 10 mm, for a total projection length **L** of between about 7 mm and about 13 mm. A crimp **116** or other suitable connection fixes the neck **114** to the wave anchor.

Each protrusion **110** folds down along the side of the wave anchor **102** when compressed for delivery, then springs up to extend nearly perpendicularly from the wave anchor **102** when released from the compressed state to the relaxed state. Specifically, the angle ϕ formed by the protrusion **110** and a leg of the wave anchor **102** may be between about 45° and about 135°, or, more preferably, between about 75° and 105°, e.g., about 80° or about 90°.

FIGS. 2A-2D are elevation and plan views of a single protrusion **110** formed of a single piece of nitinol wire with a diameter of about 0.020 inch. The wire is bent to form a pair of struts **124** that can be crimped, bonded, or welded onto a single-wire leg of an anchor (e.g., wave anchor **102** in FIGS. 1A-1D) such that the single wire of the anchor leg nestles between the struts **124**. The wire is bent to form the narrow, straight neck **114** and coiled twice to create the loop **120**. The two loops of coil form a broad, blunt edge **120** that can engage and erode the luminal wall such that the loop **112** eventually

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penetrates the luminal wall. In this case, the loops also form a face **122** that defines a plane perpendicular to the long axis of the struts **124**. When affixed to an anchor and implanted in a lumen, the face **122** is perpendicular to the long axis of the lumen and parallel to a cross section of the lumen. Alternatively, the loop **112** may be formed such that the face **122** is parallel to the long axis of the struts **124**. In this alternative orientation, the face **122** is near parallel to the lumen's long axis and near perpendicular to the lumen's cross section when implanted.

FIGS. 3A-3C illustrate how the implant **100** is secured within a lumen by a protrusion **110** with a relatively straight neck **114**. First, the implant **100** is inserted into the lumen in a compressed state, with the projection **110** folded against the collapsed anchor **102**. Once released into the lumen, the anchor **102** and the protrusion **110** expand toward their respective relaxed states, causing the edge **124** of the loop **112** to form a tent **309** in the luminal wall **301**, which may include a muscle layer, as shown in FIG. 3A. Without being bound by a particular theory, initial studies suggest that, over time, the tent **309** stretches and the face **124** erodes at the point of contact **303**, as shown in FIG. 3B. Eventually, the loop **112** erodes completely through the luminal wall **301**, as shown in FIG. 3C. Within two to four weeks, fibrotic tissue **305** forms about and through the loop **112**, securing the loop **112** with respect to the luminal wall **301**, and may secure the loop **112** in a permanent or quasi-permanent fashion (e.g., for months or years). A loop **112** that is secured in a pocket of fibrotic tissue **112** does not appear to provoke the tissue remodeling that eventually forces other projections, such as sharp protrusions, out of the intestine.

FIGS. 4A-4C show open-loop protrusions of different shapes. FIG. 4A shows a protrusion **820** formed by bending wire into the shape of the Greek letter omega, Ω . An open, straight neck **822** connects the protrusion's loop **824** to a collapsible member **102**. FIG. 4B shows a protrusion **830** formed by twisting wire into a loop. The twist forms the protrusion's neck **432** and the loop forms the protrusion's loop **834**. FIG. 4C shows a protrusion **840** with a neck **842** covered by a seal **843**. The seal **843**, which may be made of a fluoropolymer, isolates the loop **844** from the anchor **102**, and may prevent irritants from exiting the luminal wall via the channel formed by the protrusion **840**. The seal **843** does not cover the loop **844**, so tissue may grow through the loop **844**. The protrusions **820**, **830**, and **840** shown in FIGS. 4A-4C may be separate pieces of wire bonded to an anchor or they may be formed of the same piece of wire that forms the anchor.

FIGS. 4D and 4E show an open-tip protrusion **850** with an erodible section **855** that forms part of a loop **854** connected to an anchor with a straight neck. After the protrusion **850** penetrates the luminal wall, inflammatory or fibrotic tissue grows through the loop **864**, securing the protrusion **850** in the intestinal wall. Meanwhile, the erodible section **855** dissolves, turning the loop **854** into an open prong that can be removed from tissue without tearing the tissue (not shown) that forms in the opening of the loop **854**. Typically, the erodible section **855** is designed to dissolve during treatment, e.g., over six months, one year, two years, or possibly longer.

FIGS. 4F and 4G show an open-tip protrusion **860** with a loop **864** that can be detached from an anchor **102** with a narrow coupling neck **862**. As above, inflammatory or fibrotic tissue (not shown) envelops the loop **864**, securing the protrusion **860** in the intestinal wall. By disconnecting the anchor **102** from the protrusion **860**, the anchor **102** can be withdrawn endoscopically without tearing or ripping the tissue

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that envelops the loop. In some cases, the detached anchor **102** may be reinserted into the intestine and re-attached to the protrusion **860**.

FIGS. 4H-4O show alternative straight protrusions with open heads. FIGS. 4H and 4I shows protrusions **900** and **910**, respectively, with open loops **902** and **912** that define planes parallel to the lumen wall when deployed. The open loop **902** of FIG. 4H is made of a wire loop that is concentric about an axis of a straight neck **904**; whereas, the open head **912** of FIG. 4I is formed of a wire loop that is tangent to an axis of a straight neck **914**. FIG. 4K shows a straight protrusion **930** with plural open loops **932** distributed along the length of a straight protrusion **934** that extends from a collapsible anchor **102**.

FIG. 4J shows a protrusion **920** with a corkscrew-like open head **922** perched atop a straight neck **924** coupled to an anchor **102**. When deployed, the long axis of the open head **922** is roughly parallel to the long axis of the lumen, but can also be oriented perpendicular to the long axis of the lumen. Similarly, FIG. 4N shows a protrusion **960** with a whisk-shaped open head **962** at the end of a straight neck **964**. Tissue may grow about and through the openings between the windings in both the corkscrew-like head **922** and the whisk-shaped head **962**, just as in the helix protrusions described in greater detail below.

FIGS. 4L and 4M show bidirectional protrusions **940** and **950**, respectively, each of which is coupled to a collapsible anchor **102**. Each protrusion **940**, **950** includes a coil-like open head **942**, **952** that engages the luminal wall as the protrusion **940**, **950** expands from its collapsed state.

FIG. 4O shows an alternative protrusion **970** formed of a paddle with one or more openings **972**, each of which is about 0.016 inch wide. Tissue can form through the openings **972** to secure the protrusion **970** within the lumen.

The open heads may also be connected to the anchor with a detachable or erodible feature. FIG. 4P shows a protrusion **980** that includes a coiled open loop **982** connected to a straight neck **984** with a bio-erodible element **986**. Upon deployment, the loop **982** erodes through the luminal wall and soon becomes encased in fibrotic tissue, securing the protrusion **980** and attached anchor **102** in place. Over time, the bio-erodible element **986** dissolves, causing the loop **982** to become detached from the protrusion **980**. Once the head **982** is no longer connected to the protrusion **980**, the protrusion **980** can be withdrawn without necessarily tearing the scar tissue encapsulating the head **982**, making for easier removal of the implant.

Helical Protrusions with Open Loops

Alternatively, the implant may include a helical protrusion instead of a straight protrusion. The helical protrusion acts as a coil spring that pushes the open loop into the lumen wall, but in a manner that distributes the load from the collapsible anchor to the contacting tissue over a longer length as compared to a straight protrusion of similar height. Upon initial engagement with the duodenal wall, the helix, if so designed, compresses. As the tissue and helix protrusion come to equilibrium the helix approaches full expansion, causing the loop to penetrate the luminal wall. Eventually, fibrotic tissue encapsulates the loop and the expanded helix, creating a pocket that holds the loop and helix securely. Like straight protrusions with open loops, helical protrusions with open heads may be designed for permanent, quasi-permanent, or temporary implantation.

FIGS. 5A-5C are perspective views of implants that include projections with helical protrusion: FIG. 5A shows basic helical protrusions; FIG. 5B shows helical protrusions with retaining loops; and FIG. 5C shows helical protrusions

that include retaining loops and short end effects that promote initial penetration of the open loop into the muscle wall. FIG. 5A shows an implant 400 that includes five basic helical protrusions 410, each of which is coupled to a wave anchor 102 with a respective crimp 416. (Alternatively, the protrusions 410 may be sutured or releasably coupled to the anchor 102.) Each helical protrusion 410 includes a helix 414 formed of several wire coils and terminates in a loop 412 formed of two loops of wire. The opening of each head 412 is parallel to the lumen defined by the wave anchor 102. Each helical protrusion 410 has a tapered profile, with the top coils (i.e., those farthest away from the wave anchor 102) being substantially smaller than the base coils (i.e., those closest to the wave anchor 102). Each coil in the helix 414 limits the penetration of the coil above it.

The top coils are sized to focus the force from the expanding implant 400 to penetrate the duodenal wall and to ultimately elicit the healing response. Top coils approximately 3 mm in diameter are small enough to start to burrow through the muscle layer. The base coils are larger than the top coils and are sized to substantially match and blend to the crowns (vertices) of the wave anchor 102. For example, a 7 mm diameter base coil blends well to the wave anchor 102 approximately 6 mm below the crowns, but larger base coils could be used for other attachment configurations and/or anchor configurations. Typically, the outer diameter of the largest coil in the helix 414 is within the range of about 1.5 mm to about 12 mm, and the coils have an inner diameter that ranges from about 1.0 mm to about 10 mm. The loop 412 can have an inner diameter within a range of about 1.0 mm and about 6.0 mm.

The spacing of the coils or wire wraps in the helix 414 influences the tissue response. If the coils are too close together, then tissue may not be able to grow around the wire or between the coils. If the coils are too far apart, then each coil may exert more localized force on the tissue, causing the tissue to erode at the point of contact. In addition, increasing the coil spacing makes it more likely that the upper coils will infiltrate surrounding organs. Setting the spacing between wraps, or coil pitch, within a range of about 1.0 mm to about 4.0 mm (or, more preferably, within a range of about 2.4 mm to about 2.5 mm), limits the erosion caused by the upper coils while allowing for tissue encapsulation of helix 414.

In the examples shown in FIGS. 5A-5C, the loop 412 is formed of two coils of uniform diameter that are stacked upon each other, approximating a solid cylinder that does not compress. Because the loop 412 is relatively incompressible, it erodes through the duodenal wall, but only to an extent determined by the length and compliance of the helix 414. A helix 414 with appropriate compliance typically prevents the loop 412 from penetrating much beyond the muscle layer of the duodenal wall.

FIG. 5B shows an implant 430 with several projections 440, each of which includes a retrieval element 442 that extends from the loop 412 towards the wave anchor 102. In the example shown in FIG. 5B, the retrieval element 442 is a loop of wire formed with an optional hypodermic tube 472, which provides an additional surface for fibrotic tissue to encapsulate; this further encapsulation may increase the anchoring strength. Each retrieval element 442 fits in the conical cavity defined by its associated helical neck 414 and can be used to exert a force normal to the axis of the conical cavity on the helical protrusion 410. For example, a normal force can be used to prevent or slow expansion of the helical protrusion 410 or to collapse an expanded helical protrusion 440. Other retrieval elements may be hooks, balls, or other suitable features for applying a normal force to the helix.

In the example shown in FIG. 5B, a constraint 444 threaded through the retrieval element 442 keeps the helical protrusion 440 in a fully or partially collapsed state. In some cases, the constraint 444 is a suture or biodegradable element that allows the user to influence how quickly the helical protrusion 440 expands after implantation, which, in turn, affects how quickly the loop 412 penetrates the luminal wall. Releasing tension on the suture or engineering the decay time of the biodegradable element allows the helix 414 to open to its full height more slowly, prolonging the equilibrium time and slowing the effect of the helical protrusion 440 on the contacting tissue.

A drawstring (not shown) that runs through some or all of retrieval elements 442 can be used to withdraw the protrusions 440 from the luminal wall. Pulling on the drawstring applies a normal force directly to the loops 412, causing the loops 412 to collapse into the coils below to disengage the helix 414 from the surrounding tissue. As the coils collapse, one within the next, they act as a “cheese cutter”: each coil helps to shear the surrounding tissue from the coil above it as the above coil passes through the lower coil, freeing the helical protrusion 440 from any scar tissue that may have grown through or around the wire in the loop 412 and the helix 414. Pulling on the drawstring also causes the anchor 102 to collapse for endoscopic withdrawal from the implantation site as described below.

FIG. 5C shows an implant 460 with an end effect 462 at the end of each helical protrusion 470. In this example, each end effect 462 is a post that is oriented in the center of a respective helical protrusion 470 and protrudes slightly beyond the loop 412 of the respective helical protrusion 470. The end effects 462, which may be sharpened to engage the contacting tissue more quickly, initiate an injury to the duodenal wall and lead the heads 412 through the duodenal wall. Because each end effect 462 pierces the luminal wall, it initiates the injury that causes fibrotic tissue to encapsulate its associated helical protrusion 470 more quickly. Quickly embedding the helical protrusion 470 is important for stabilizing and maintaining placement of the implant in a mobile vessel with pressurized luminal contents, such as the duodenum or intestines.

FIG. 6 is a perspective view of an alternative helical protrusion 510 with a retrieval element 542 formed of a single wire without a hypodermic tube. The wire is coiled to form a helix 514 and a loop 512, then folded and formed into a retrieval element 542. Excess wire extending from the tail of the retrieval element 542 is trimmed and may be sharpened to create an end effect 562. The base coil of the helix 414 can be trimmed and/or bent as necessary before the projection is attached to the wave anchor 102, e.g., with a crimp 416, as shown in FIGS. 5A-5C, or using any other suitable attachment. Alternatively, the helical protrusion can be fabricated with a post that runs up its center, and the post can be crimped or otherwise affixed to the anchor 102.

FIGS. 7A-7C show how the helical protrusion 410 shown in FIG. 5A engages a luminal wall to secure an implant 400 within the lumen. The implant 400 is inserted into the lumen in a compressed state, with the helical protrusion 410 collapsed against the collapsed anchor 102, as shown in FIG. 7A. Releasing the helical neck 414 allows the helical neck 414 to expand, causing a tent 603 to form in the duodenal wall 601, as shown in FIG. 7B. As the neck 414 continues to expand against the duodenal wall 601, it pushes the loop 412 through wall 601, as shown in FIG. 7C. Scar tissue 607 forms about and possibly through the loop 412 and neck 414. Without being bound by any particular theory, initial studies suggest that helical necks 414 tend to encourage more fibrotic encap-

sulation than straight necks of similar height because helical necks have more wire in contact with the tissue.

The compliance of the helical neck **414** affects how quickly the loop **412** penetrates the luminal wall **601**. Initial studies suggest that the top-most coils in the helical neck **414** continue to push through tissue after initial contact until the contacting tissue and helix **414** come to equilibrium. If the helical neck **414** is as compliant as the luminal wall, however, then the neck **414** will not be able to push the loop **412** through the luminal wall **601**. Since the compliance of the helical neck **414** is largely a function of wire diameter and pitch, increasing either the wire diameter or the pitch the wire diameter generally increases the rigidity of helical neck **414**. Increasing the wire diameter too much may make it difficult to form the wire into tight loops to shape the loop **412**. Wire with a diameter in the range of about 0.016 inch to about 0.040 inch is generally suitable for helical protrusions **410**. Nitinol wire with a diameter of about 0.019" offers a balance: it can be formed into tight bends for the end of the helical neck **414** and the loop **412**, yet forms a helix that is stiffer than the luminal wall **601**. It can also be packed into a capsule for endoscopic delivery. The diameter of the helix **414** can also be varied to further customize the transition in stiffness and tissue response.

Although FIGS. 5-7 show a helix **414** with a linear transition between successive coil diameters, alternative helices can have other shapes, including parabolic profiles, cylindrical profiles, hourglass profiles, and conical profiles (e.g., with the vertex of the cone connected to the anchor). Alternatively, the helix **414** can be formed in a flattened coil that is narrow at the center and flares out from the center into a flat spiral shape. The helix **414** could also be formed of a post that terminates in a coil with its wrappings aligned or angled with respect to one another in a corkscrew-like fashion. Compared to other three-dimensional shapes, tapered shapes tend to be easier to disengage from a mating surface. Parabolic shapes transition more quickly from large coils to small coils, facilitating a lower profile protrusion. Similarly, transitions between coils or wraps in the helix **414** can be customized as desired. For example, the coils in the helix **414** can be sized such that each coil fits into the coil below. This sizing of successive coils facilitates a lower profile for packing into the delivery catheter and facilitates disengagement from the duodenal wall.

Compliance Measurements

The compliance/stiffness of the protrusions disclosed herein can be characterized, in part, by the force required to deflect the protrusions from their respective relaxed (extended) states towards their respective collapsed states. For a protrusion with a straight neck (e.g., protrusion **110** of FIGS. 2A-2D), compliance may be defined, in part, by the normal force required to deflect the protrusion at room temperature by a given amount towards the strut of the collapsible anchor. Measurement shows that applying a force of at least about 0.1 lbf normal to the head (i.e., parallel to the long axis of the lumen) completely collapses a straight-necked protrusion made of 0.010-inch diameter nitinol wire, with a total length of 13 mm, ending in a loop formed of two wraps of wire with an inner diameter of about 3 mm. Similar measurement shows that applying about 0.8 lbf normal to the head deflects the head by about 0.250 inch for a straight-necked protrusion made of 0.020-inch diameter nitinol wire, with a total length of 11.5 mm, ending in a loop formed of two wraps of wire with an inner diameter of about 3 mm. Other straight-necked protrusions may be deflected by about 0.250 inch from their relaxed positions by forces within a range of about 0.80 lbf to about 0.95 lbf.

The compliance of a helical protrusion can be characterized, in part, by measuring the force required to (partially) collapse the helical protrusion at room temperature. Measurement shows that applying a force normal to the long axis of a helical protrusion within a range of about 0.19 lbf to about 1.75 lbf, or, more preferably, about 0.32 lbf to about 0.95 lbf, collapses the protrusion by about 0.250 inch, depending on the wire diameter, coil pitch, and coil size:

TABLE 1

Normal force applied to compress nitinol helical protrusions by 0.250 inch at room temperature

Protrusion Height	Base Coil Diameter	Top Coil Diameter	Coil Spacing	Wire Diameter	Normal Force
10 mm	6 mm	3 mm	2.4 mm	0.016"	0.19 lbf
6 mm	6 mm	3 mm	4.0 mm	0.023"	0.32 lbf
10 mm	6 mm	3 mm	2.4 mm	0.028"	0.95 lbf
10 mm	6 mm	3 mm	2.4 mm	0.030"	1.75 lbf

In addition to the compliance of the helix as measured in the normal force to compress the helix, resistance to bending must be considered. Helix stiffness can also be characterized by the force required to deflect the helix sideways, i.e., in the plane normal to the long axis of the helix. A balance must be struck between compressability and rigidity. Deflecting a nitinol helical protrusion with a 6 mm height, 6 mm base coil diameter, 3 mm top coil diameter, 4.0 mm coil spacing, and 0.020-inch wire diameter sideways by 0.250 inch at room temperature takes a force of at least about 0.033 lbf. Increasing the wire diameter to about 0.028 inch increases the force to about 0.135 lbf for a 0.250-inch deflection at room temperature. A preferred balance can be defined within the specifications above.

Deployment and Removal of Anchors Secured with Loops and Necks

Each of the aforementioned implants may be deployed in the intestine, preferably in the duodenum, and more preferably in the duodenal bulb just distal to the pylorus. Typically, a doctor or other qualified person inserts the implant into the intestine with an endoscopic delivery device. During insertion, the delivery device holds the implant in a compressed state. Once in position, the implant is released from the delivery device and allowed to self-expand, causing each neck coupled to the anchor to push its respective loop against the intestinal wall. Some implants may include a sleeve coupled to the anchor, which can be deployed within the intestine as described in U.S. Pat. Nos. 7,122,058; 7,329,285; 7,678,068; and U.S. patent application Ser. No. 11/057,861, filed on Feb. 14, 2005, by Levine et al., all of which are incorporated herein by reference in their entireties.

An implant secured with protrusions tipped with open loops may be removed laparoscopically, surgically, or, more preferably, endoscopically with an endoscope.

For example, an implant may be collapsed using a draw-string, then withdrawn from the intestine using an endoscope. Further details on endoscopic removal can be found in U.S. application Ser. No. 11/318,083, filed on Dec. 22, 2005, by Lampert and Melanson; and in U.S. application Ser. No. 12/005,049, filed on Dec. 20, 2007, by Levine et al., both of which are incorporated herein by reference in their entireties. Seals, Sleeves, and Restrictor Plates

FIG. 8 shows an implant **700** that includes an anchor **702** with a polymer covering **704**. Protrusions **710** projecting from the anchor **702** support open loops **712** that can be used to create fibrotic encapsulations in the intestinal wall as

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described above. A sleeve **706** is coupled to the distal side of the anchor **702** for extension into the intestine. The sleeve **706** may be permanently or detachably affixed to the anchor **702**. For instance, a detachable sleeve can be endoscopically attached to or removed from a permanently or semi-permanently secured anchor depending on treatment progress.

Typically, the sleeve **706** is floppy and conformable to the wall of the intestine when deployed. It also has a wall thickness of less than about 0.001 inch to about 0.005 inch and a coefficient of friction of about 0.2 or less. The polymer covering **704** and the sleeve **706** may be made of a fluoropolymer, such as ePTFE coated or impregnated with fluorinated ethylene polyethylene (FEP), or any other suitable material. The sleeve **706** and anchor covering **704** can be a single, integrally formed piece. They can also be separate pieces, depending on whether the anchor **702** is partially or wholly uncovered, as long as the anchor **702** forms a sufficiently good seal between the sleeve **706** and the stomach, pylorus, and/or intestine to funnel chyme through the sleeve **706**. Each loop **712** remains uncovered or only partially covered to promote the in-growth of fibrotic tissue.

Anchors secured with loops and necks may also be used to secure restrictor plates within the gastrointestinal tract to treat obesity, such as the restrictor plates disclosed in U.S. patent application Ser. No. 10/811,293, filed on Mar. 26, 2004, by Levine et al.; U.S. patent application Ser. No. 11/330,705, filed on Jan. 11, 2006, by Levine et al.; and U.S. patent application Ser. No. 11/827,674, filed on Jul. 12, 2007, by Levine et al., all of which are incorporated herein by reference in their entireties. An implant with a restrictor plate typically includes a restricting aperture that retards the outflow of food from the stomach to the intestine. The diameter of the aperture is less than 10 mm, is preferably less than 7 mm, and is more preferably initially in the range of about 3-5 mm. Alternatively, the aperture may be elastic and expandable under pressure from material flowing through the anchor and the aperture at elevated physiological pressures; as pressure increases, the aperture opens to greater diameters. The implant may include a sleeve that extends into the intestine.

While this invention has been particularly shown and described with references to example embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the scope of the invention encompassed by the appended claims. For example, all or part of the protrusions described above can be covered to further control interaction with contacting tissue. A bio-absorbable suture or adhesive could be used to affix the covering to the protrusion. As the bio-absorbable material degrades or is absorbed by the body, the covering is free to fan open, creating an added level of control of interaction between the protrusion and the surrounding tissue. Alternatively, the protrusion may be made from a polymer or a composite material, such as a non-degradable or biodegradable material. Implants can also include different types of protrusions, e.g., any combination of straight protrusions with open loops, helical protrusions with open loops, and even pointed barbs.

What is claimed is:

1. An implant comprising:

- a collapsible anchor configured to be deployed within a lumen of a gastrointestinal tract, the collapsible anchor having a relaxed diameter of at least 40 mm; and
- a protrusion having a first end connected to the collapsible anchor and a second end formed in a loop defining an opening for tissue ingrowth, the protrusion having a length of at least 5 mm, the protrusion including an extension that connects the loop to the collapsible

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anchor, the protrusion expanding from a collapsed state, in which the extension having the loop is collapsed alongside the anchor, to a relaxed state, in which the protrusion extends outward from the anchor upon deployment within the gastrointestinal tract, the loop having a blunt edge to engage a wall of the lumen, and the protrusion configured to, upon deployment, cause the blunt edge of the loop to form a tent in the wall of the lumen, the tent, over time, stretching the wall of the lumen and the blunt edge eroding the wall of the lumen at a point of contact, fibrotic tissue forming about the loop to secure the loop with respect to the wall of the lumen.

2. The implant of claim 1 wherein the protrusion includes a straight neck that connects the loop to the collapsible anchor.

3. The implant of claim 1 wherein the protrusion includes a helix that connects the loop to the collapsible anchor.

4. The implant of claim 3 wherein the helix has a tapered profile.

5. The implant of claim 3 further including a constraining element configured to maintain the helix in a collapsed state.

6. The implant of claim 3 further including:
an end effect at a tip of the loop.

7. The implant of claim 1 wherein the loop defines an opening with an inner width within a range of 1 mm to 13 mm.

8. The implant of claim 1 wherein the loop defines an opening with an inner diameter of 3 mm.

9. The implant of claim 1 wherein the protrusion extends between 6 mm and 13 mm from the collapsible anchor upon full deployment from the collapsible anchor.

10. The implant of claim 1 wherein the loop is formed of wire with a diameter of 0.010 inch to 0.040 inch.

11. The implant of claim 1 wherein, in a relaxed state, the protrusion extends outwards from the collapsible anchor at an angle of between 45 degrees and 135 degrees.

12. The implant of claim 11 wherein, in a relaxed state, the protrusion extends outwards from the collapsible anchor at an angle of 80 degrees.

13. The implant of claim 1 wherein the loop defines a plane that is perpendicular to the wall of the lumen.

14. The implant of claim 1 wherein the loop defines a plane that is parallel to the wall of the lumen.

15. The implant of claim 1 wherein the loop is formed of a wire loop at an end of the protrusion.

16. The implant of claim 1 further including:
a drawstring configured to collapse the protrusion towards the collapsible anchor to disengage the protrusion from the tissue.

17. The implant of claim 1 further including additional protrusions connected to the collapsible anchor, each additional protrusion having an end formed in a loop.

18. The implant of claim 1 wherein the collapsible anchor is a wave anchor.

19. The implant claim 1 further including:
an unsupported, thin-walled sleeve coupled to the collapsible anchor and configured to extend into the lumen upon deployment of the collapsible anchor.

20. The implant of claim 1, wherein fibrotic tissue forms about and through the loop in a range of 2-4 weeks.

21. A method of securing a collapsible anchor within a lumen of a gastrointestinal tract, the method comprising:
deploying the collapsible anchor within the lumen, the collapsible anchor including a protrusion having a first end coupled to the collapsible anchor and a second end formed in a loop defining an opening for tissue ingrowth, the protrusion including an extension that connects the

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loop to the collapsible anchor, the protrusion expanding from a collapsed state, in which the extension having the loop is collapsed alongside the anchor, to a relaxed state, in which the protrusion extends outward from the anchor upon deployment within the gastrointestinal tract, the loop having a blunt edge to engage a wall of the lumen; and

causing the blunt edge of the loop to form a tent in the wall of the lumen, the tent, over time, stretching the wall of the lumen and the blunt edge eroding the wall of the lumen at a point of contact, fibrotic tissue forming about the loop to secure the loop with respect to the wall of the lumen.

22. The method of claim 21 wherein the collapsible anchor has additional protrusions attached thereto, each additional protrusion having a loop, and further including:

allowing the loops of at least some of the additional protrusions to penetrate a wall of the lumen.

23. The method of claim 21 wherein deploying the collapsible anchor includes allowing the protrusion to bend away from the collapsible anchor to an angle of between 45 degrees and 135 degrees.

24. The method of claim 21 wherein deploying the collapsible anchor includes allowing the protrusion to bend away from the collapsible anchor to an angle of 80 degrees.

25. The method of claim 21 wherein penetrating the wall of the lumen with the loop includes engaging the wall with the edge of the loop.

26. The method of claim 21 wherein the protrusion includes a straight neck that connects the loop to the collapsible anchor, and further including:

bending the protrusion alongside the collapsible anchor to place the implant in a collapsed state; and
inserting the implant, in the collapsed state, into the lumen.

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27. The method of claim 21 wherein the protrusion includes wire formed in a helix that connects the loop to the collapsible anchor, and further including:

inserting the helix, in a collapsed state, into the lumen; and
releasing the helix from the collapsed state.

28. The method of claim 27 wherein penetrating the wall of the lumen further includes allowing fibrotic tissue to grow between coils in the helix.

29. The method of claim 27 further including:

collapsing the helix with a drawstring to disengage the helix from the tissue; and

withdrawing the collapsible anchor from the lumen.

30. The method of claim 29 wherein collapsing the helix further includes shearing fibrotic tissue with coils of the helix.

31. The method of claim 21 wherein penetrating the wall of the lumen with the loop includes engaging the wall with a face of the loop.

32. The method of claim 21 wherein penetrating the wall of the lumen with the loop includes engaging the wall with an end effect at a tip of the loop.

33. The method of claim 21 further including:

extending an unsupported, thin-walled sleeve coupled to the collapsible anchor into the lumen.

34. The implant of claim 16, wherein the drawstring runs through either the opening in the loop or an additional retaining loop connected to the protrusion.

35. The implant of claim 29, wherein the drawstring runs through either the opening in the loop or an additional retaining loop connected to the protrusion.

36. The method of claim 21, wherein fibrotic tissue forms about and through the loop in a range of 2-4 weeks.

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